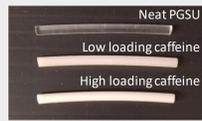


PURPOSE

Commonly used materials for long-acting implantables (LAI) comprise non-biodegradable polymers that rely on diffusion-mediated drug release. These LAIs lack the ability to achieve controlled release kinetics for multi-month therapies that require high drug loadings (>40 wt%). This affects both water-soluble drugs, where rapid diffusion occurs, and water-insoluble drugs, where slow diffusion leads to insufficient release. While biodegradable polymers exhibit benefits over non-biodegradables, bulk-degrading biodegradables similarly suffer from the inability to achieve steady release at high loadings. Surface-eroding biodegradables such as Hydralsee™ (PGSU) (poly(glycerol sebacate) urethane) (Secant Group), a synthetic biodegradable elastomer for controlled drug release,¹ offer a superior polymeric delivery system able to provide zero-order release at high drug loadings over many months. Using caffeine as a model drug, *in vitro* and *in vivo* studies demonstrate Hydralsee (PGSU) implants exhibit minimal burst release followed quickly by steady-state release kinetics.



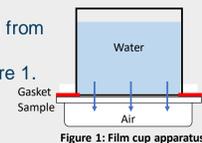
OBJECTIVE(S)

Demonstrate the effectiveness of surface-eroding Hydralsee (PGSU) loaded at high percentages of caffeine to maintain controlled release kinetics over many months using *in vitro* and *in vivo* analyses, as well as its manufacturability using a solvent-free room temperature extrusion technology.

METHOD(S)

Formulation

- Target Hydralsee (PGSU) crosslink density selected using inverted film cup method adapted from ASTM E96.
- Various loaded-Hydralsee (PGSU) formulations were tested using apparatus depicted in Figure 1.
- Depth of water penetration through matrix determined using SEM after four weeks at 37.0°C.

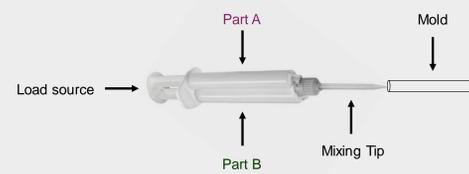
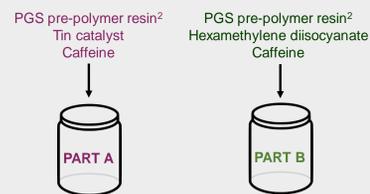


Caffeine-loaded Hydralsee (PGSU) Implant Manufacture

- Hydralsee (PGSU) implants were synthesized using a two-part chemistry paired with dual-barrel syringe extrusion technology.
- Methodology was optimized to eliminate common obstacles like air pockets, uneven mixing, and short working time.
- Rods were filled into molds and complete crosslinking of the urethane polymer was achieved at room temperature.

Step 1: Materials combined separately using dual-asymmetric centrifugal mixing

Step 2: Two parts combined and extruded using dual-barrel syringe fitted with a static mixing tip into mold



Benchmark, *In Vitro* and *In Vivo* Analyses

- Final 0, 40, and 60 wt% caffeine-loaded rods were analyzed for caffeine dispersion, content uniformity, crosslink density, compressive modulus, and chemical functionality using SEM, HPLC, TGA, MTS, and FTIR, respectively.
- In vitro* release was monitored using a flow-through cell dissolution apparatus under physiological pH and temperature with 0.1M phosphate buffered saline.
- HPLC was used to quantify caffeine concentration in weekly collected media to generate a release profile over a two-month period (study still underway).
- In vivo* release was assessed using a subcutaneous implantation rat model and LCMS to generate pharmacokinetic profiles from weekly plasma collections over two months (study still underway).
- Histology was performed on explanted tissue to evaluate Hydralsee (PGSU) biocompatibility and degradation.

RESULT(S)

Target Crosslink Density Determined by Film Cup Experiments

- Cross-sectional SEM images of 60 wt% caffeine-loaded Hydralsee (PGSU) after film cup analysis demonstrate Hydralsee's (PGSU) ability to reduce water permeation by increasing crosslink density.
- Additionally, permeation is reduced in a highly loaded Hydralsee (PGSU) film containing another hydrophilic API that does not exhibit agglomerations as is commonly seen with caffeine.
- Penetration of water into a polymer/drug matrix is a direct factor in burst release and dose-dependent release kinetics. Therefore, it can be extrapolated that high crosslink density will reduce burst release and achieve steady release kinetics *in vitro* and *in vivo*. This is a direct result of Hydralsee (PGSU) alone exhibiting impermeability and only 2-3% swelling in water.



Homogeneous Rod Implants Successfully Formed Using Dual-barrel Syringe Method

- Resultant crosslink density, content uniformity, and compressive moduli were consistent across a representative 40% and 60% caffeine-loaded Hydralsee (PGSU) rod batch (Figures 2 and 3).
- Scanning electron microscopy images depicted good dispersion of the caffeine within the Hydralsee (PGSU) matrix.
- Within multiple rod batches, tight selection criteria were applied to implants for their use *in vitro* and *in vivo*.

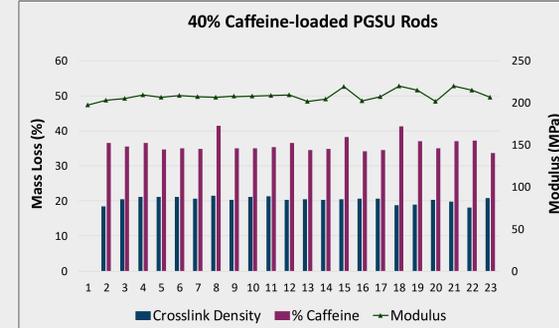


Figure 2: Crosslink density, % caffeine, and moduli results for a representative 40% caffeine-loaded Hydralsee (PGSU) rod batch

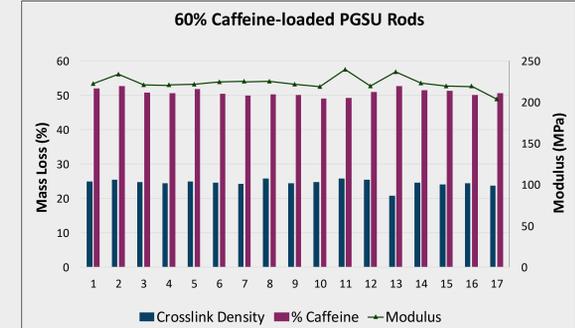


Figure 3: Crosslink density, % caffeine, and moduli results for a representative 60% caffeine-loaded Hydralsee (PGSU) rod batch

In vitro Release Kinetics of Highly Loaded Hydralsee (PGSU)

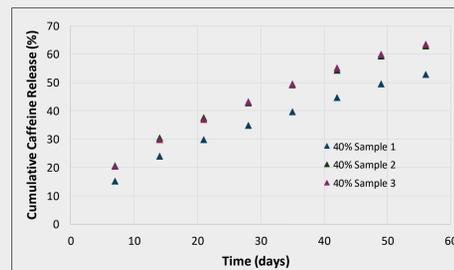


Figure 4: *In vitro* release kinetics of 40% caffeine-loaded Hydralsee (PGSU) rods quantified by LCMS

- No lag observed in reaching zero-order release from onset through two-month time period with minimal burst release.
- Data demonstrates the ability of Hydralsee (PGSU) to resist water permeation and swelling, generating a true surface erosion-dependent release mechanism.
- Release rate is consistent across the manufactured rod batch.

In Vitro and *In Vivo* Higuchi Modeling

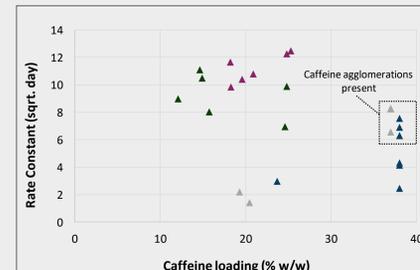


Figure 5: Higuchi modeling comparison between *in vitro*, *in vivo* high crosslinked rods and *in vitro*, *in vivo* low crosslinked rods

- Highly crosslinked rods show similar rate constants across 20-40% loadings, demonstrating release rate is independent of caffeine content.
- Calculated rate constants for high crosslinked rods are significantly lower than is commonly observed for EVA, PCL, and PLGA.

In vivo Release Kinetics and Histology of Highly Loaded Hydralsee (PGSU)

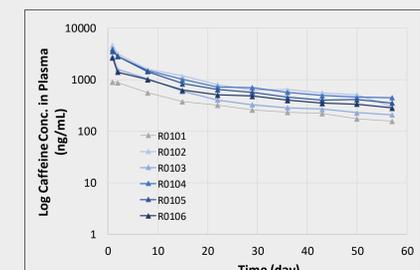


Figure 6: Pharmacokinetic curve of 40% caffeine-loaded Hydralsee (PGSU) rod implants in rats

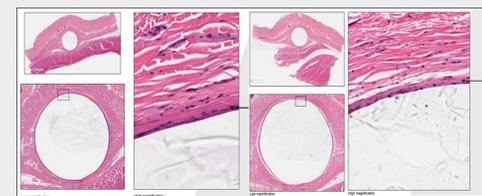


Figure 7: Histology staining on control animal (left) and animal containing drug-loaded implant (right). Arrows indicate minimal fibroplasia.

- Highly loaded Hydralsee (PGSU) showed minimal burst release *in vivo* and sustained zero-order kinetics for two months.
- Expected duration of delivery 6-10 months based on clearance rate of 45 mL/hr.³
- Release rates consistent across the six implants.
- Histology three months post-implantation showed little to no inflammation or fibroplasia for all animals.
- No macrophages, lymphocytes, or fibrous encapsulation observed in both control and drug-loaded Hydralsee (PGSU).

CONCLUSION(S)

For numerous treatments being studied today, LAIs capable of delivering a steady dose over months to years are needed. Current materials on the market rely on diffusion-based release kinetics, whether degradable or non-degradable, and therefore have difficulty being loaded at high percentages and still exhibit zero-order release for multiple months. Hydralsee (PGSU) behaves as a true surface-eroder demonstrated *in vitro* and *in vivo*, and paired with its low swellability and water permeability, can be loaded at high concentrations and still exhibit low burst release and zero-order release. Furthermore, challenges in manufacturability regarding API and crosslink density homogeneity have been overcome in conjunction with the development of reliable analytical techniques. Ultimately, Hydralsee (PGSU) implants are an attractive carrier providing efficient pharmacokinetics, longer-lasting treatment at higher API loadings, and enhanced patient comfort via improved implant flexibility, when compared to traditional polymers.

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